

***Smeringopina ebolowa* Huber, 2013**

Huber, B. A. 2013. Revision and cladistic analysis of the Guineo-Congolian spider genus *Smeringopina* Kraus (Araneae, Pholcidae). *Zootaxa* 3713: 1-160.

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***Smeringopina ebolowa* new species**

Figs. 6, 628–632, 678–679, 694, 716–737

Smeringopina Cam61: Dimitrov, Astrin & Huber 2013 (DNA data).

Type. ♂ holotype from Cameroon, South Region, near Ebolowa (2°54.9'N, 11°08.3'E), 620 m a.s.l., near ground, 11.–12.iv.2009 (B.A. Huber), in ZFMK (Ar 10282).

Other material examined. CAMEROON: *South Region*: near Ebolowa, same data as holotype, 8♂6♀ in ZFMK (Ar 10283); same data, 4♀ 10 juvs. in pure ethanol, in ZFMK (Cam 102). Near Ebolowa, Nkoetye (2°51.4'N, 11°21.7'E), 700 m a.s.l., near ground, 12.iv.2009 (B.A. & J.C. Huber), 5♂3♀ in ZFMK (Ar 10284); same data, 1♂1♀ 1 juv. in pure ethanol, in ZFMK (Cam 142). Near Kribi (2°54.0'N, 9°54.4'E), 20 m a.s.l., near ground, 9.iv.2009 (B.A. Huber), 4♂13♀ in ZFMK (Ar 10285); same data, 2♀ 5 juvs. in pure ethanol, in ZFMK (Cam 133).

Assigned tentatively: CAMEROON: *Centre Region*: forest fragment in Yaoundé (3°48.8'N, 11°32.6'E), ~700 m a.s.l., under large log, 13.iv.2009 (B.A. & J.C. Huber), 2♂ in ZFMK (Ar 10286); same data, 2 juvs. in pure ethanol, in ZFMK (Cam 99). Yaoundé, Zenker leg., no further data, 1♂ in ZMB. Yaoundé, Mt. Febe, 900 m a.s.l., 21.ii.1981 (R. Bosmans), 2♀ 2 juvs. in MRAC (167985). Near Yaoundé, Mt. Kala (3°51.0'N, 11°20.3'E), 730 m a.s.l., near ground, 14.iv.2009 (B.A. Huber), 1♂1♀ in ZFMK (Ar 10287); same data, 1♀ 2 juvs. in pure ethanol, in ZFMK (Cam 103). *Littoral Region*: near Edéa, Koukoué (3°41.2'N, 10°06.4'E), 50 m a.s.l., near ground, 8.iv.2009 (B.A. & J.C. Huber), 2♂1♀ 2 juvs. in ZFMK (Ar 10288); same data, 2♀ 7 juvs. in pure ethanol, in ZFMK (Cam 112). Unidentified and unknown localities: Cameroon, Ercalera(?), E. Simon coll. number 22016, no further data, 1♂1♀ in MNHN (Ar 10470). Cameroon, no further locality data, 9.vi.1939 (J. Félix), 1♂ in MNHN.

Etymology. The name is a noun in apposition, derived from the type locality.

Diagnosis. Easily distinguished from most known congeners by modified male clypeus (slightly projecting rim set with modified (cone-shaped) hairs: Figs. 725, 731); from species with similar clypeus (*S. simintang*, *S. kribi*, *S. bwiti*) by shape of procurus (massive, with short distal apophysis, without ventral branch, Figs. 716–717); also by relatively large modified hairs on male chelicerae (Fig. 722; similar only in *S. mayebout*), and parallel ventral ridges on procurus prolaterally (visible only after detachment of bulb; Fig. 716). Female not easily distinguished from similar species with roughly triangular anterior epigynal plate evenly curved in lateral view (especially *S. belinga*).

Male (holotype). Total body length 6.8, carapace width 1.7. Leg 1: 78.0 (17.6 + 0.7 + 17.7 + 38.7 + 3.3), tibia

2: 12.1, tibia 3: 8.4, tibia 4: 10.9; tibia 1 L/d: 100. Distance PME-PME 170 μm , diameter PME 170 μm , distance PME-ALE 90 μm , distance AME-AME 35 μm , diameter AME 160 μm . Carapace ochre-yellow with brown mark posteriorly and brown lateral margins; ocular area brown posteriorly, clypeus with pair of dark marks in lower half, sternum dark brown; legs ochre-yellow, dark rings subdistally on femora and tibiae and in patella area, tips of femora and tibiae whitish; abdomen ochre-gray with distinct black pattern dorsally, laterally, and ventrally. Habitus as in Figs. 628–629, ocular area slightly elevated, secondary eyes with distinct ‘pseudo-lenses’; clypeus slightly projecting, with about 12 modified (cone-shaped) hairs near rim (Figs. 725, 731); deep thoracic pit and pair of shallow furrows diverging behind pit (Fig. 726). Chelicerae as in Figs. 722 and 732, with lateral proximal apophyses and large distal apophyses, the latter and frontal cheliceral face provided with very large modified (cone-shaped) hairs (Figs. 733–735). Palps as in Figs. 630–632; coxa unmodified; trochanter with simple retrolatero-ventral apophysis; femur proximally with ventral pocket bordered retrolaterally by strong sclerotized ridge (Fig. 727), with small retrolateral apophysis, without prolateral projection; prolateral femur-patella joint very prominent and strongly shifted toward ventrally (hidden by bulb in Fig. 630); tarsus with some stronger hairs dorsally (Fig. 730); procurus with distinct hinge between proximal and distal part, with distinctive ridges prolaterally (visible only after detachment of bulb; Fig. 716); bulb with heavily sclerotized proximal part of embolus (Figs. 718, 728). Legs without spines and curved hairs, with few vertical hairs, retrolateral trichobothrium on tibia 1 at 1%; prolateral trichobothrium present on all tibiae; pseudosegments barely visible. Gonopore with two epiandrous spigots (Fig. 736).

Variation. Males from South Region all very similar to holotype, but with slight variation in number of modified hairs on male chelicerae and clypeus (up to about 20) and in size and distinctiveness of ridges and teeth on sclerotized proximal part of embolus. Males from Centre Region and Littoral Region show additional slight differences and are therefore assigned tentatively: shapes of ventro-distal apophyses on procurus slightly different (retrolateral apophysis shorter and less pointed; prolateral apophysis longer); dorso-distal sclerotized part of procurus either stronger (Yaoundé area) or less developed (Koukoué); prolateral ridges of procurus fewer but stronger in males from Yaoundé area; modified hairs near median line of male chelicerae more widely spaced in males from Yaoundé area; entire procurus either slightly more slender (Koukoué) or shorter (“Ercalera”); cheliceral apophyses more slender in males from Yaoundé area. Even more pronounced variation occurs in sclerotized proximal part of embolus (Figs. 719–721). Tibia 1 in 15 other males (all localities combined): 13.9–18.3 (mean 16.4).

Female. In general similar to male but clypeus unmodified; clypeus variably dark; posterior median light line ventrally on abdomen absent. Tibia 1 in 22 females: 10.8–14.0 (mean 12.9). Epigynum large, consisting of evenly curved (in lateral view), roughly triangular anterior plate and large posterior plate (Figs. 678–679, 723, 737); internal genitalia as in Figs. 694 and 724.

Natural history. *S. ebolowa* was found in well preserved forests (e.g. in a small canyon near Ebolowa) and in disturbed forest patches (e.g. in man-made holes in the ground in disturbed forest fragments near Koukoué and Nkoetye). The large webs were built in sheltered spaces near the ground.

Distribution. Known from several localities in southwestern Cameroon (Fig. 627).



6. *S. ebolowa*, male (Kribi, Cameroon).

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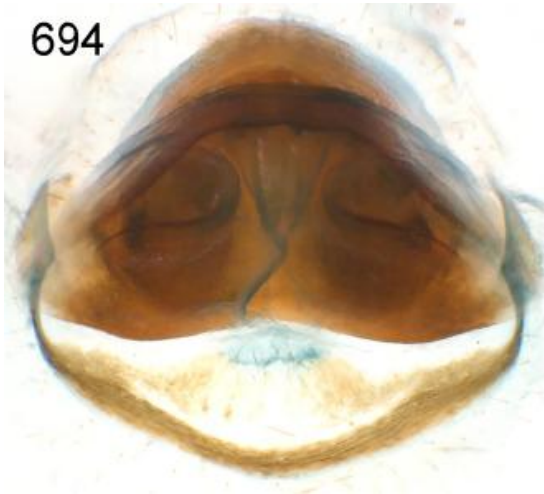
FIGURES 628–637. *Smeringopina ebolowa* n. sp. (628–632) 628–629, 633–634. Males, dorsal, lateral, and ventral views. 630–632, 635–637. Left male palps, prolateral, dorsal, and retrolateral views. Arrow points at ventral sclerotized ridge.

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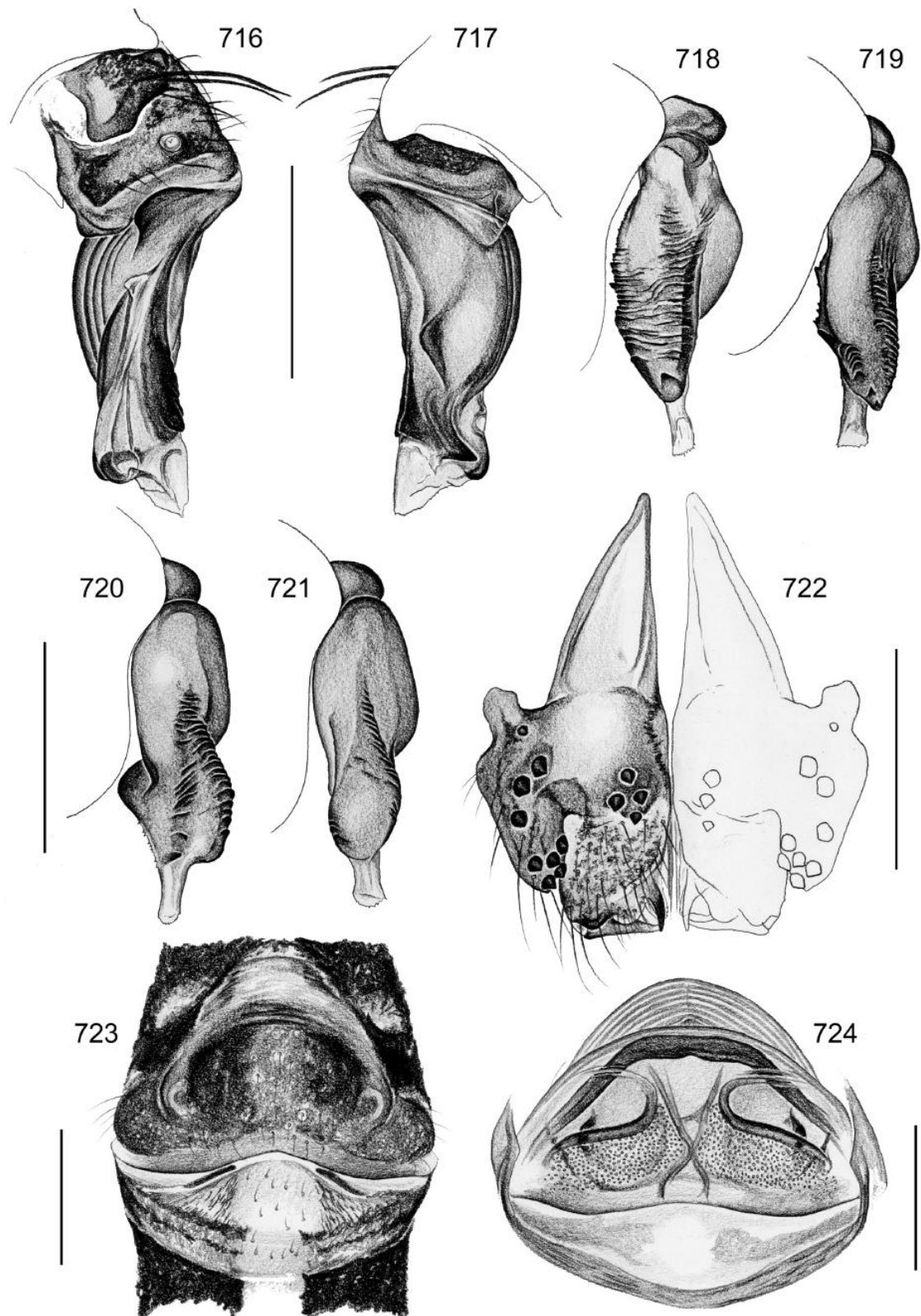


678–679. *Smeringopina ebolowa* n. sp.

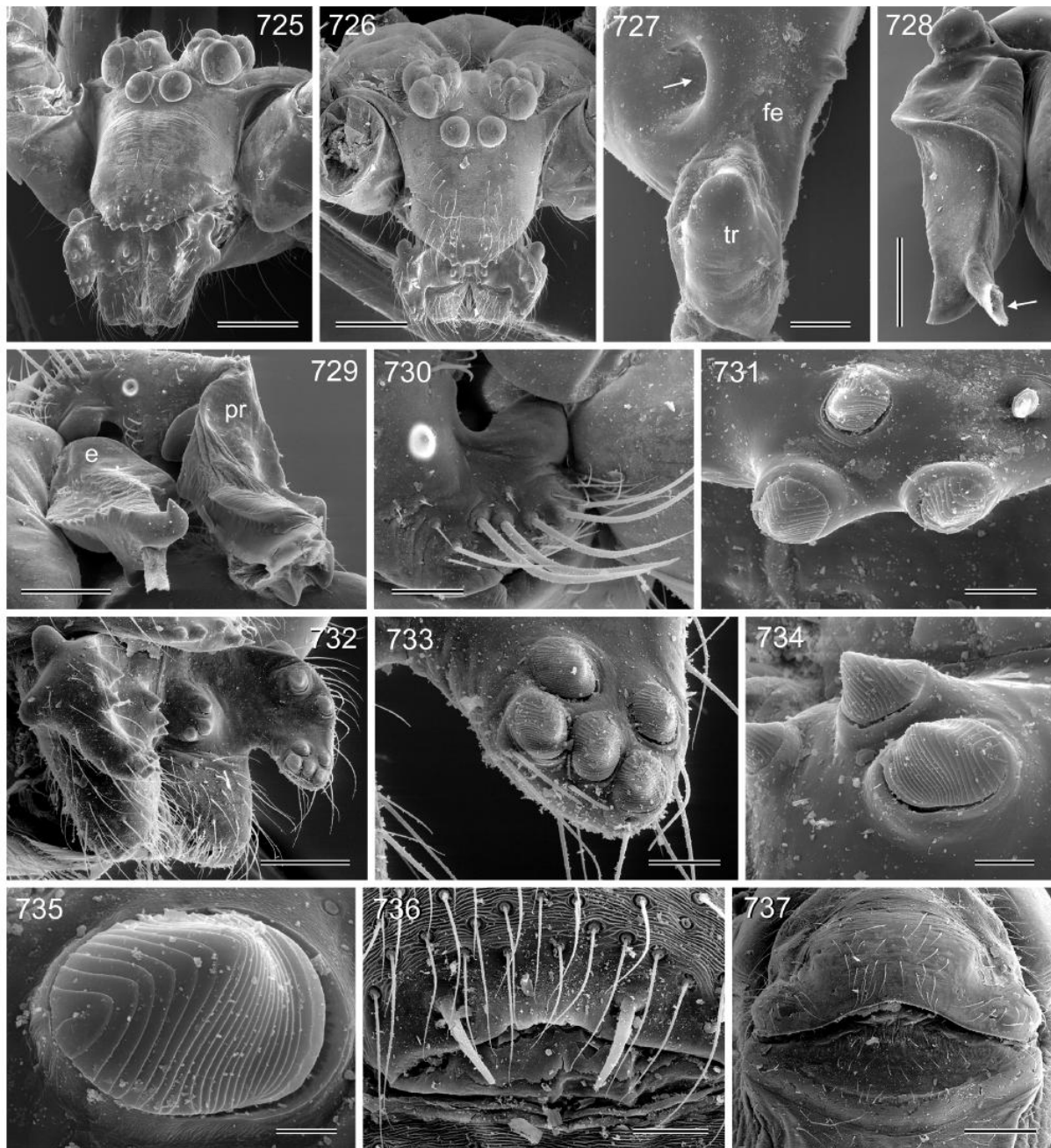
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694. *Smeringopina ebolowa* n. sp.



FIGURES 716–724. *Smeringopina ebolowa* n. sp. 716–717. Left procurus, prolateral and retrolateral views (type locality). 718–721. Left emboli, prolateral views, males from near Ebolowa (type locality, 718), near Kribi (719), Yaoundé (720), and Koukoué (721). 722. Male chelicerae, frontal view (type locality). 723. Epigynum, ventral view. 724. Cleared female genitalia, dorsal view. Scale lines: 0.5 (all emboli at same scale).



FIGURES 725–737. *Smeringopina ebolowa* n. sp. 725–726. Male prosoma, oblique frontal and fronto–dorsal views. 727. Left palpal trochanter and femur, ventral view; arrow points at ventral pocket. 728. Embolus; arrow points at sperm duct opening. 729. Left procurus and embolus, prolatero–distal view. 730. Macrotrichia and tarsal organ of left male palp. 731. Modified hairs on male clypeus. 732. Male chelicerae. 733. Tip of left male cheliceral apophysis. 734–735. Modified hairs on male chelicerae. 736. Male gonopore. 737. Epigynum, ventral view. Abbreviations: e: embolus; fe: femur; pr: procurus; tr: trochanter. Scale lines: 10 μm (735), 20 μm (734), 30 μm (731), 40 μm (733), 50 μm (736), 80 μm (730), 100 μm (727), 200 μm (728–729, 732), 300 μm (737), 400 μm (725–726).